

# Translation Enhancement of m-Rna by Immuno Evasion Proteins

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**Abstract**— In this paper, mono-functional EKB i.e. “E3”, “K3” and “B18R” derived from vaccinia virus is compared with multi-functional NS1 derived from “influenza A virus” to compare their ability for the enhancement of the mRNA translation. Further, expression of luciferase was transferred by delivery of unmodified luciferase assorted with NS1 mRNA and compared with delivery of unmodified luciferase assorted with either “E3”, “K3” or “B18R” respectively. Various combination of “E3”, “K3”, “B18R” were assorted with NS1-TX91 mRNA at different ratios and delivered with m RNA of luciferase.

**Keywords:** luciferase, NS1 (non-structural protein 1), EKB, m-RNA, translation

## I. INTRODUCTION

Recent clinical trials and preclinical studies have shown excellent potential for mRNA therapy. “Encoding antigens”, “transcription factors”, “growth factors”, “in-vitro-transcribed” (IVT) mRNA can be used as a cancer and infectious disease vaccine or as a “protein replacement therapy”, “genome editing” or “cell reprogramming [1].

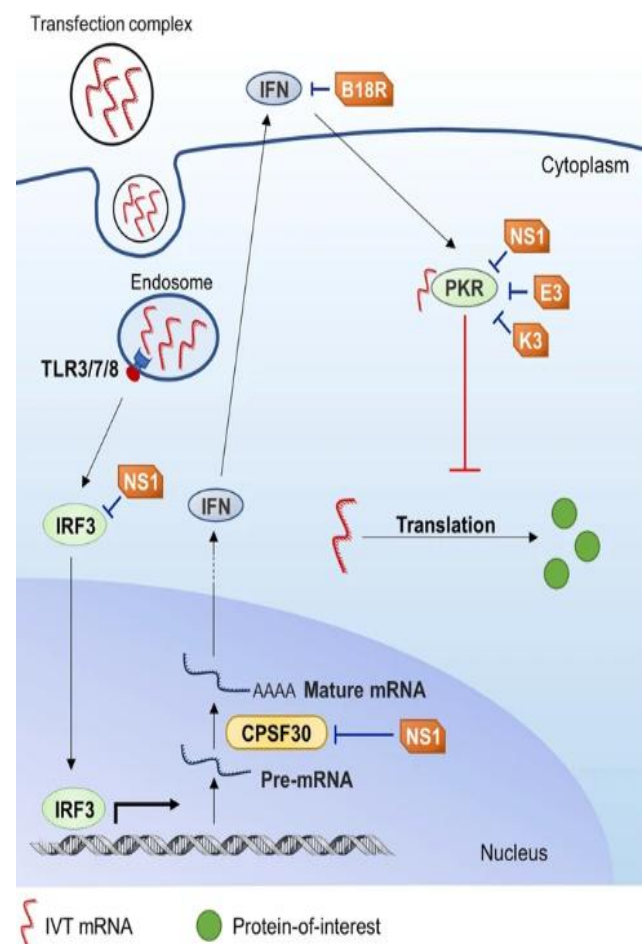
In- vitro transcribed m-RNA can induce negative response against viruses by stimulating “endosomal” and “pattern recognition receptors” (PRRs), including TLRs (toll like receptors) that includes TLR 3, TLR 8, TLR 7, “protein kinase RNA” (PKR), “retinoic acid-inducible gene I” (RIG-I), and “2’-5’ oligoadenylate synthetase” (OAS)[2].

In order to improve mRNA translation, an immune evasion approach using viral immune evasion proteins has been suggested. A plethora of “immune evasion proteins” have developed in nature to thwart host antiviral reactions upon viral infection.

By co-delivering “immune evasion proteins” and mRNA-of-interest encoding mRNA, immune reactions caused through transfection are efficiently suppressed and translation of m-RNA-of-interest is significantly improved [3]. “EKB” and “NSP1” have been reported to enhance translation of m-RNA of interest.

E3 and K3 inhibit PKR activation, and B18R obstructs “type I IFN signaling” by seizing binding of extracellular IFN to “IFN $\alpha/\beta$  receptor” (IFNAR) on the membrane of the cell [4]. NS1 is rather multifunctional in contrast. It inhibits immune-related- proteins such as OAS, PKR, NF- $\kappa$ B, “interferon regulatory factor 3” (IRF3) as well as “non-immune-related- proteins” such as “cleavage and factor 30 (CPSF30) polyadenylation”[5].

Binding and inhibiting CPSF30 by NS1 results in accumulation of nonprocessed pre-m-RNA within the nucleus and impedes the expression of host genes, including hundreds of IFN-stimulated genes (ISGs)[6], [7].



**Figure 1: working mechanism of EKB and NS1 in inducing innate immune response**

In this research, “EKB” and NS1 with subtype TX91 that is extracted from “strain A / Texas/36/91” were compared on the basis of their capacity to improve mRNA translation. The verification for the effectiveness of these “immune response protein” was done. NS-1 was found more substantial than EKB. Due to lack of supremacy between “NS1-TX9” and EKB, and release of PR8, supremacy of NS1 multifunctionality was established.

## II. MATERIALS

“Human foreskin fibroblast cell line” that includes “BJ fibroblasts”, “ATCC”; “Human hepatocellular carcinoma cell line” that includes “HepG2”, “ATCC”; fetal bovine serum (FBS) ;penicillin-streptomycin 100 $\times$  solution; and trypsin 0.5% 10 $\times$  solution; Alamar Blue stock solution ; transfection kit of Stemfect mRNA; “GLO lysis buffer”; “GLO luciferase reagent”.

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III. METHOD

Cloning of EKB genes

Synthesis of EKB genes was carried out by “integrated DNA technologies” (IDT). PCR amplification was done for synthesised genes and further cloned into “Sal-1” and “Not-1” sites of a vector (pGEM4Z-A64).

In-vitro transcription

Plasmids without B18R gene, were transformed into a linear form with Spe-1. Further it was purified and was used as a template for “in-vitro transcription” by using “T7 High Yield RNA Synthesis Kit” in the presence of an “anti-reverse cap analog” (ARCA) to produce capped transcripts with approximately 80 percent capping efficiency. Capped transcripts were further used to produce m-RNA modified with pseudouridine. B18R gene containing plasmid were transformed into a linear form with Sac-1. Transformed plasmids were purified and finally transcribed. Polyadenylation of the transcript was performed with *E.coli* “poly (A) polymerase” (PAP) and confirmed by performing “gel electrophoresis”. RNeasy kit was used to purify synthesized m-RNA. Spectrophotometer analysis was carried out for quantification and confirmation of full-synthesized m-RNA was done by “agarose gel electrophoresis” [8].

In-vitro transfection

“Stemfect RNA transfection kit” was used for transfection study [9]. “BJ fibroblast” and “HepG2” were placed on a 96 micro-titer plate with a cell density of “ $1.2 \times 10^4$ ” and was incubated for night. For pseudo modified m-RNA pretreatment, cells were introduced into 40 nanogram of pseudomodified m-RNA. After 6 hours, the fresh medium was added to replace the previous medium and transfected with 40 nanogram of unmodified “luciferase (Luc)” m-RNA. After 18 hours, “Alamar Blue assay” and “Luc assay” were outperformed.

Cells were transfected with “40 nanogram per well of Luc m-RNA and EKB, GFP m-RNA or NS1-TX91 to determine the co-delivery of “immune evasion genes” [10]. To determine the synergism between NS1-TX91 and EKB, transfection of the cells were carried out with Luc (20 ng) and NS1-TX91 (20 ng) alone or in combination of NS1-TX91 with “E3”, “K3” and/or B18R.

To determine synergism between “E3”, “K3”, PR8 variants, transfection of the cells were carried out with Luc (20 ng) and PR8 mutants (20 ng) alone or in combination with E3, K3, green fluorescent protein (GFP) [11].

Biochemical assay

Further biochemical assay was carried out to determine the cell viability, and quantify the luciferase expression.

IV. RESULTS

Translation enhancement

In this paper it was seen that NS1-TX91 could enhance the m-RNA translation more effectively as compared to EKB.

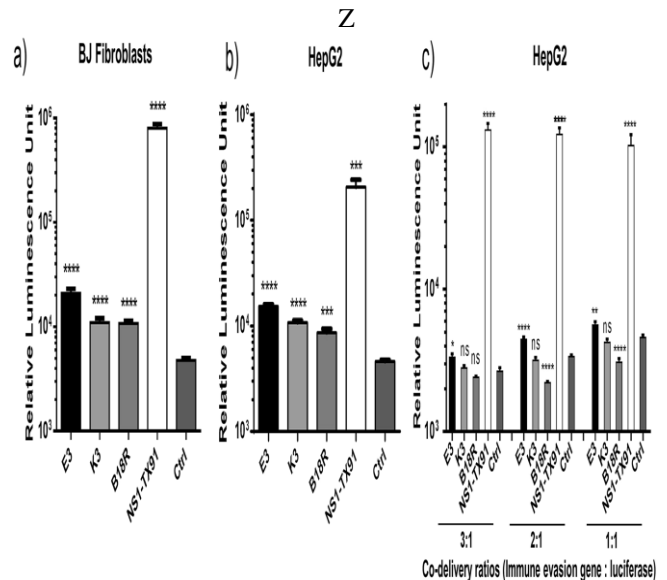


Figure 2: comparison of EKB and NS1-TX91 in translation enhancement

Figure 2a: comparison of EKB and NS1-TX91 with BJ Fibroblasts, Figure 2b: comparison of EKB and NS1-TX91 with HepG2 cells, Figure 2c: comparison of EKB and NS1-TX91 with HepG2 cells for co-delivery “immune responsive protein”.

Figure 2 a,b shows that, m-RNA encoding numerous “immune responsive protein” demonstrated higher m-RNA translation of luciferase as compared to “green fluorescent protein” control.

Figure 2c, demonstrated that, NS1-TX91 showed a 10 fold increase in luciferase production than EKB at different ratios.

Interferon suppression by m-RNA transfection

Suppression of interferon production was assayed after 18 hours by collecting supernatant of HepG2, and BJ fibroblast and IFN-beta and transfecting with “EKB”, “GFP” or “TX91”. Further it was measured by using technique ELISA [12]. It was concluded that “EKB” and “TX91” suppressed immune response more effectively at the time of transfection.

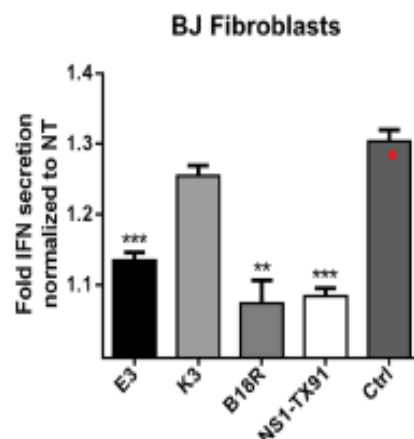


Figure 3a: interferon suppression in BJ Fibroblasts



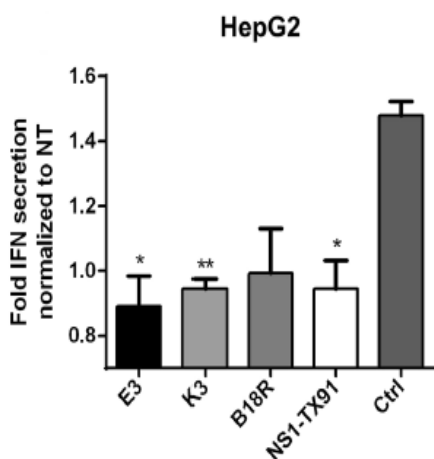


Figure 3b: interferon suppression in HepG2

Host gene expression inhibition

“Host gene expression inhibition” (HGEI) expression was verified by the co-delivery of HGEI with plasmid luciferase (mutants). Table 1 depicts the gain of “host gene expression inhibition” an loss of PKR inhibition.

Abbreviation	Amino acid positions						IRF3	CPSF30	PKR	Ref
	38	41	103	106	123	124	R38, K41	F103, M106	I123, M124	
PR8 <sup>WT</sup> (PR8 <sup>C+/+</sup> )	R	K	S	I	I	M	Yes	No	Yes	11
PR8 <sup>IRF3-</sup>	A	A	S	I	I	M	No	No	Yes	11
PR8 <sup>PKR-</sup> (PR8 <sup>C-/-</sup> )	R	K	S	I	A	A	Yes	No	No	11
PR8 <sup>C+/-</sup>	R	K	F	M	A	A	Yes	Yes	No	—
PR8 <sup>C+/+</sup>	R	K	F	M	I	M	Yes	Yes	Yes	11

Table 1: immune evasion process for HGEI and PKR inhibition

V. CONCLUSION

The potential of viral “immune evasion proteins” i.e. “E3”, “K3”, “B18R” and “NS1-TX91” was verified to enhance translation of m-RNA. It was concluded that “NS1-TX91” showed characteristic performance in “BJ fibroblast” and “HepG2” as compared to “EKB”. Synergism was also not seen between “NS1-TX91” and “EKB”. Finally, “EKB” and “NS1” showed translation enhancement but, NS1 showed super performance in enhancing m-RNA translation.

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