
Puttamade Gowda. J, Prasanna Kumar S C

Abstract: White blood cells (leucocytes) are main constituents of blood, plays a major role in immune system. They rescuing our body from foreign materials and infectious diseases. All five types of WBCs are origin from multipotent cell bone marrow also called hematopoietic stem cells. These five types are grouped into granulocytes and agranulocytes. Leukemia a blood cancer occur due to abnormal development of leukocytes (WBCs). In united states 62130 people receive leukemia diagnosis in 2017 and 245000 deaths occurs by this disease its effects are even more in undeveloped countries. Counting and identifying of white cells in images of microscope is so tedious, time taking and the results depends on experience of hematologist. The proposed algorithm is a combination of spectral-spatial method for segmenting cytoplasm and nucleus of WBCs from the microscopic, spectral and hyperspectral images. Here we use an Integrated method of improved spectral angle mapper (ISAM) with Gram-Schmidt orthogonalization process and k-means segmentation process. The resulting spatial spectral information of WBCs gives an accuracy of above 95% for both nuclei and cytoplasm.

Keywords: white blood cell, improved spectral angle mapper, orthonormal set

I. INTRODUCTION

Analysis of microscopic blood cells is a powerful tool of diagnose the blood related diseases. Among three main constituents of blood cells, Leukocytes/nucleus. Its diameter range is 6µm to 20µm, and they play major part in immune condition of the body. They fight against cancer cells and other toxic materials. Hematologist set a valuable information by identifying and inspecting the WBCs in peripheral blood smear. So it is easy to diagnose the disease like blood cancer i.e. leukemia and such diseases of WBC. But blood study manually under microscope is a delayed, subjective and tedious, results is mainly based on the skill of the technician and his experience.

Rapid development in the field of information science, tends to automate of WBC analysis depends on digital image processing, identification of pattern approaches. This computer-aided method is used in pointing out, and computation of white cells of cytological blood images.

In fast one to one and half decades many researcher can develop different algorithm for segment and identify different parameters about WBCs from microscopic blood smear images.

II. RELATED WORK

Many research works are going on segmentation and detection of WBC from blood images. In Hayan T madhloom et.al [1] work localization and segmentation of lymphoblast by morphological reconstruction process. Subrajeet mohapatra et.al [2] propose image segmentation using shadowed C-means(SCM) clustering method. They compare the tested result with other classifier like Navie Bayesian, KNN, MLP, RBIN and SVM.

Qing Li et.al[3] separate the cytoplasm and nucleus of WBC from spectral and hyperspectral images using pixel-wise ISAM segmentation then they apply majority voting method for within snakes regions.

Doriniet.al [4], uses three different methods for separating cytoplasm and nucleus of WBCs. To remove background they use bottom–hat transform, flood-fill operation for remove RBCs and for extracting segmented nucleus they use Watershed transform algorithm. For extracting the same Baidya Nath shah et.al[5] apply PCA followed by active contour (snake) algorithm.

In S.Mohapatra et.al [6], for obtaining WBC they use a fuzzy based color segmentation process, use contour signature and Hausdorff dimension for classifying nucleus. In our work[7] for extracting nucleus we use a combined technic of fuzzy c with snake and get the better result than above work and also count the number of WBC in a given image.

S.H.Rezafotighi et al [8], and M. Mohamed [9] et al applies Gram-Schmidt orthogonalization algorithm for segmenting the nuclei of WBC. In our second work[10] , [11] we use the same technic and get a orthonormal image and using K-means segmentation we get nuclei of white blood cells and classify according to their characteristics. Like different researcher uses different technics for extracting WBC from its image. M.Y.Mashor et al [12], adopts global contrast stretching algorithm and use HSI color segmentation process for improving the quality of image.

M.Y.Mashor, F.U.Francis et.al [15] apply partial contrast stretching procedure for image enhancement succeeded by image segmentation. For extract WBCs features and screening the image using MLP.

M.Y.Mashor [16] applies two procedures, In their initial part enhance white blood cell region they use contract enhancement method on suspected leukemia images and in next part they apply HIS color space image segmentation process.

Monica Madhukar, Anyony.T.C et al [17] show microscopic images by exposing color correlation, contrast enhancement followed by K-means clustering method on resultant images, from this they detecting nucleus.

Fabio Scotti [18] apply Fuzzy K-means clustering on microscopic image there by eliminating unwanted images and enhance the required image of WBC.

D.C.Haung et al [20] by using Otsu’s threshold method for extracting nucleus of white cells, for texture measurement they use co-occurrence matrix. In their work they clearly differentiate five types of leucocytes by considering available features from principal component analysis (PCA).

The five types of leucocytes are Neutrophils:- very popular among all five types, counts of 50-70% of all white cells, circulatory, short lived having minute light granules. These are multi-lobed nucleus and each lobe is interconnected by a thin strands. Strands are made of nuclear material.

Eosinophils:- granulocytes cells, large granules, pink or red in color. These cells having two lobes in each nucleus, their counts are less than 5% count of leucocytes.

Basophils:- these cells having large number of granules,. When stained they appear purple or deep blue in color. Their presence is less than 1% of white cells.

Lymphocytes:- these agranular cells having well developed cytoplasm, when stained looked pale blue. The nucleus covers whole cell surrounded by a small rim of cytoplasm. Their counts are 25-35% of leucocytes.

Monocytes:- they have large nucleus, and agranular cells. Nucleus have ‘U’ shape or kidney been shape, they have abundant cytoplasm of light blue in color and their counts are 3-4% of WBCs.

III. METHODOLOGY

Fig.1 shows the architecture of proposed system, initially the image is converted into gray-scale in preprocessing, followed by noise will filtered using median filters and normalization is takes place.

A. Pre-processing

B. Gray conversion:- Processing time of RGB input color image is more compared to its equalent gray image. So it is necessary for convert RGB to gray.

Fig. 1 algorithm of segmentation of WBC nucleus and cytoplasm

Filtering:- Median filter filters every pixel in an image from its neighboring pixels. Then decides the pixel belongs to surrounding or not. This filter replaces each pixel value by median value of neighboring pixel, thereby sorting neighborhood pixels in numerical order. Middle most sorted value is taken by pixel value retained it as median value. A window will formed by neighboring pixels. Windows takes different shapes and centered around desired pixels.

A median value is calculated by

\[
me(Y) = \begin{cases} 
Y_{(m+1)} & \text{for } N = 2m + 1 \\
\frac{1}{2} (Y_{(m)} + Y_{(m+1)}) & \text{for } N = 2m 
\end{cases} \tag{1}
\]

Then normalization is done as in our previous paper. Improved Spectral Angle Mapper :-

After getting Normalized molecular spectral images of the blood smear, to determine spectral similarity of these spectrum by calculating the angle between this with reference spectrum. In spectral image each selected pixel naming as unknown spectrum ‘a’ and reference spectrum ‘b’ taken from spectral library. The spectral angle between these two spectrums is calculated by

\[
\theta(a,b) = \arccos\left(\frac{a \cdot b}{|a||b|}\right) = \arccos\left(\frac{\sum_{i=0}^{N-1} a_i b_i}{\sqrt{\sum_{i=0}^{N-1} a_i^2} \cdot \sqrt{\sum_{i=0}^{N-1} b_i^2}}\right) \tag{2}
\]
To rectifying noise there by reducing the wavelength shift error, Spectral angle calculated between test and reference spectrum, we adopt Improved Spectral Angle Mapper (ISAM) method in which, we calculate spectral angle by move backward and forward 2 bands respectively. Then select maximum angle as the spectral angle between two vectors.

Gram-Schmidt orthoganalization process:-
This process takes a linearly independent, finite set \( S=\{b_1, b_2, \ldots, b_k\} \) for \( k<n \) and generating an orthonormal set \( S'=\{d_1, d_2, \ldots, d_k\} \) both are same span and inner product space generally called Euclidean space \( \mathbb{R}^n \).

The projection operator is defined as
\[
\text{Proj}_d b = \frac{\langle d, b \rangle}{\langle d, d \rangle} \frac{d}{\langle d, d \rangle} \quad \text{(3)}
\]

Where \( \langle d, b \rangle \) ...... inner product of \( d \) & \( b \). The operator projects 'b' vector orthogonally on to 'd' vector. It works as follows
\[
\begin{align*}
  d_1 &= b_1 \\
  e_1 &= \frac{d_1}{\|d_1\|} \\
  d_2 &= b_2 - \text{proj}_{d_1} b_2 \\
  e_2 &= \frac{d_2}{\|d_2\|} \\
  d_3 &= b_3 - \text{proj}_{d_1} b_3 - \text{proj}_{d_2} b_3 e_1 = \frac{d_3}{\|d_3\|} \\
  \vdots \\
  d_n &= b_n - \sum_{j=1}^{n-1} \text{proj}_{d_j} b_n e_n = \frac{d_n}{\|d_n\|}
\end{align*}
\]

here \( d_1, d_2, \ldots, d_n \) is orthonormal vectors and \( e_1, e_2, \ldots, e_n \) forms a orthonormal set. Using this method, from linearly independent set \( S=\{b_1, b_2, \ldots, b_n\} \) we select one vector ' \( b_k \)' which is maximum orthogonal and remove all other vectors which are minimum orthogonal over N-dimensional space.

The required vector \( W_k \) is given by
\[
W_k = b_k - \sum_{j=1}^{k-1} \text{proj}_{d_j} b_k
\]

So the result ' \( W_k \)' from inner product of set ' \( S \)' is
\[
\begin{align*}
  \langle b_j, w_k \rangle &= 0 & \text{for} & 1, 2, \ldots, n \text{ and } j \neq k \\
  \langle b_k, w_k \rangle &= k & k \neq 0
\end{align*}
\]

In fig. below shows the relation between \( w_k \) & \( b_1, b_2 \) and \( b_3 \) is graphically shown in 3-D spaces

**IV. RESULTS AND DISCUSSION**

The blood smear hyperspectral or microscopic spectral images are taken as input images. Initially these images required filter and normalization. Then suitable spectral image as explained by ISAM procedure as explained above. After that select a ortho-normal vector by using Gram-Schmidt orthoganalizing procedure to above image. Then by applying K-means clustering on these orthonormal image we get a perfectly accurate cytoplasm and nucleus of white blood cells.

![a) input image](image1)

![b) grayscale image](image2)

![c) ISAM image](image3)

**Fig. 3 explanation of Gram-Schmidt process**
In the above result we can clearly separate the nucleus and cytoplasm of white blood cell of input image. The fig.3(a) is input image, fig.3(b) is its grayscale equalent fig.3(c) shows resultant image of ISAM method, fig.3(d) is an ortho-normal image selected from Gram-Schmidt orthogonalization process fig.3(e) shows the K-means clustered image whereas fig.3(f) and 3(g) shows Nucleus and cytoplasm of WBC respectively. The result can be clearly explained by Histogram shown in fig.4. Histogram of input image is shown in fig.4(a) with histogram of nucleus and cytoplasm is shown by fig.4(b) and 4(c) respectively.

Fig. 4 histogram of a) input grayscale image b) nucleus of WBC & c) cytoplasm of WBC

Fig. 3 Test image with Intermediate Results

d) ortho-normal image
c) K-means clustered image
f) Nucleus of WBC
g) Cytoplasm of WBC
Fig. 5 Result from the proposed system for different images. a- original input images. b- gray scale images. c- image from ISAM process. d- ortho-normal image selected from Gram-schmidt. e- k-means clustered image. f- Nucleus of WBC. g- Cytoplasm of WBC.

Method evaluation:-

Presence of Blast cell can be identified in input image by these steps: a) Identification of WBCs in an image. b) Lymphocyte selection. c) classified as tumor cell. The proposed benchmark has following test.

Cell test :: each single cell tested as blast or not.
Image level :: the image test is positive if we find at least one blast cell in an image.

For all Benchmark the following terms are used.

- True positive(TP) : Number of correctly classified positive element by test.
- True negative(TN) : number of correctly classified negative element by the test.
- False positive(FP, type 1 error.) : the number of element by the test classified as positive but they are not.
- False Negative(FN, type 2 error): the number of elements by the test classified as negative but they are not.

Using above definitions we process the following parameters: accuracy, Error rate, specificity, sensitivity, false positive and false negative rate. The performance evaluation is tabulated as below.

Table. 1 Performance Evaluation

<table>
<thead>
<tr>
<th>parameters</th>
<th>Formula</th>
<th>Result in %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Accuracy</td>
<td>(TN+TP)/(TN+TP+FP+FN)</td>
<td>98.38</td>
</tr>
<tr>
<td>Error rate</td>
<td>(FN+FP)/(TN+TP+FP+FN)</td>
<td>1.62</td>
</tr>
<tr>
<td>Sensitivity</td>
<td>TP/(TN+TP)</td>
<td>96.23</td>
</tr>
<tr>
<td>Specificity</td>
<td>TN/(FP+TN)</td>
<td>98.10</td>
</tr>
<tr>
<td>False Negative rate</td>
<td>FNR=FN/(TP+FN)</td>
<td>3.25</td>
</tr>
<tr>
<td>False positive rate</td>
<td>FPR=FP/(TN+FP)</td>
<td>2.15</td>
</tr>
</tbody>
</table>
V. CONCLUSION

For diagnose the disease in medical field segmentation of WBC plays a vital role. It is common test to know the condition of WBC of the patient for provide him a proper treatment. Segmentation manually requires more resources and the results depends on the experience of the hematologist. So these automated segmentation will proposed in this Improved Spectral Angle Mapper, Gram-Schmidt orthogonalization, K-means segmentation process are used to get the spatial and spectral characteristics of the images by that we get a more better result than previous work.

REFERENCES

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